



Baseline

Mercury levels in selected bycatch fish species from industrial shrimp-trawl fishery in the SE Gulf of California

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ABSTRACT

Baseline Hg concentration in bycatch fish from the SE Gulf of California were determined in muscle and liver of 19 species. Levels of Hg in muscle were compared with legal limits of this element in national and international legislation. Considering all fish species, mean concentrations in liver ($2.458 \pm 1.997 \mu\text{g g}^{-1}$) were significantly higher ($p < 0.05$) than in muscle ($0.993 \pm 0.670 \mu\text{g g}^{-1}$). The sequence of averaged Hg concentrations in most ichthyofauna was liver > muscle. Highest level of Hg in muscle ($2.556 \mu\text{g g}^{-1}$) and liver ($7.515 \mu\text{g g}^{-1}$) corresponded to *Diapterus peruvianus* and *Ophioscion strabo*, respectively. Considering muscle samples, none of the species had levels of Hg above the limit ($1.0 \mu\text{g g}^{-1}$ wet weight) in the Mexican legislation; with respect to the Japanese ($0.4 \mu\text{g g}^{-1}$ wet weight) and British ($0.3 \mu\text{g g}^{-1}$ wet weight) legislations, 26.3% and 31.6% of the species respectively, were above the corresponding limits.

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In Mexican waters, specifically in the Gulf of California, the shrimp trawl fishery is the main source of bycatch, a total of 114,000 t of fish, crustaceans and mollusks (equivalent to a total biomass of $90 \pm 45 \times 10^3$ t) are discarded every year (Madrid-Vera et al., 2007). Among bycatch species, fish is the most abundant group (Rábago-Quiroz et al., 2008); the families mainly represented (as number of species) as bycatch (López-Martínez et al., 2001) are Sciaenidae (34), Paralichthyidae (18), Haemulidae (16) and Carangidae (16). The number of fish species (>200) in the Gulf of California is relatively high in comparison with ichthyofauna from other areas in the Mexican Pacific (Amezcua-Linares, 1990); the overlapping of geographical distribution of fish from contiguous zoogeographic provinces results in an elevated biodiversity in this part of the Eastern Pacific Ocean (Mora and Robertson, 2005). Considering the elevated biomass of fish species captured as bycatch, diverse alternatives to use these fish have been proposed; for example in Venezuela, diverse bycatch fish were used to prepare hamburgers, breaded fillet, pasta and salami (Cabello et al., 2005), Madrid-Vera et al. (2007) suggest that in Mexico most of the fish caught as bycatch could be filleted and thus give it an increased value.

Most of the captured fish inhabit coastal zones and it is known that elevated discharges of industrial and urban wastes near to the

shores are important sources of trace metals. These contaminants are considered as relevant from the ecological and toxicological point of view. Mercury (Hg) is one of the most dangerous elements in the marine environment; it is supplied to the environment from natural (mineral deposits, volcanoes, forest fires and oceanic emissions) and anthropogenic (cement kilns, roasting of sulfide ores for acid production and smelting for gold, copper, iron, lead and zinc production) sources (Hylander and Meili, 2003). The issue of Hg occurrence in fish is of major environmental concern, in fact, accumulation of Hg in biota is the most important features of Hg cycle in the marine environment (Fitzgerald et al., 2007). To our knowledge, no information related to Hg levels in bycatch fish from the Gulf of California has been published; considering the above statement, the main objectives of the present study were: (a) to determine baseline Hg concentration in muscle and liver of bycatch fish, and (b) to compare Hg levels in the edible portion of fish with legal limits of this element in national and international legislation.

Fish were obtained during March 2011 from the shrimp trawl fishery that operates at the continental shelf in the SE Gulf of California at a depth range of 30–46 m (Fig. 1). Fish species were selected considering their importance as a food source for humans. In the laboratory, most fish were identified to species level; after total length and weight were recorded, specimens were dissected. Muscle tissue from the median dorsal area and liver were used for the analysis; samples were frozen at -20°C . Glassware and plastic utensils were previously washed according to Moody and

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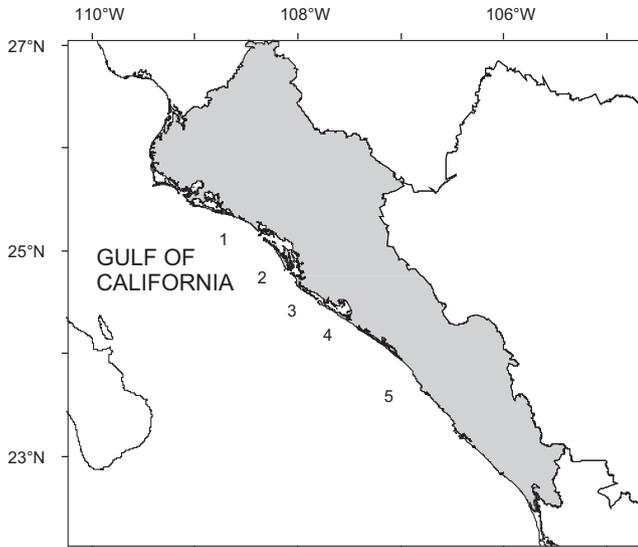


Fig. 1. Sites of shrimp trawling where bycatch fish were collected.

Lindstrom (1977). Samples were freeze-dried for 72 h ($-52\text{ }^{\circ}\text{C}$ and 60×10^{-3} mbar) in a Labconco Freeze-dry System-FreeZone 6, then ground in an agate mortar with pestle (Fischer-Scientific). Powdered samples (0.25 g) were acid digested (5 ml of concentrated nitric acid-trace metal grade, Baker) using capped teflon vials (Savillex™) on a hot plate (Barnstead Thermolyne) during 3 h ($120\text{ }^{\circ}\text{C}$). Digested samples were stored in polyethylene containers for further analysis. Analyses of Hg were made by cold vapor atomic absorption spectrophotometry (CV-AAS) in a mercury analyzer (Buck Scientific-model 410) (UNEP, 1993). Validation of the analytical process was assessed by analyzing certified reference materials (DORM-3, fish protein; DOLT-4, dogfish liver). Blanks and reference materials were run with every batch of 20 samples. Results are reported in $\mu\text{g g}^{-1}$ on a dry weight basis, detection limit was estimated in $0.012\text{ }\mu\text{g g}^{-1}$ dry weight. Recoveries for DORM-3 and DOLT-4 reference materials were 90% and 102%, respectively. Since legal limits of Hg are given on a fresh weight basis, conversion of Hg concentrations from dry weight (Hg_{dw}) to fresh weight (Hg_{fw}) was made according to the equation: $\text{Hg}_{\text{fw}} = \text{Hg}_{\text{dw}} * (100 - \% \text{ humidity}) / 100$ (Magalhães et al., 2007). Differences of Hg levels with the feeding habit of fish and comparison of Hg concentrations between muscle and liver of all fish species were made by a Stu-

Table 2

Concentrations of Hg ($\mu\text{g g}^{-1}$ dry weight) in selected tissues of studied ichthyofauna from the state of Sinaloa (SE Gulf of California).

Species	Muscle	Liver
<i>Larimus argenteus</i>	0.900 ± 0.752	4.622 ± 4.585
<i>Haemulopsis axillaris</i>	1.696 ± 1.022	4.358 ± 2.668
<i>Micropogonias ectenes</i>	0.656 ± 0.701	1.078 ± 0.791
<i>Selar crumenophthalmus</i>	0.555 ± 0.496	4.820 ± 2.899
<i>Trachinotus kennedyi</i>	0.106 ± 0.080	0.528 ± 0.646
<i>Diapterus peruvianus</i>	2.556 ± 1.171	3.186 ± 2.947
<i>Cyclosetta querna</i>	0.394 ± 0.236	0.536 ± 0.532
<i>Epinephelus acanthistius</i>	0.238 ± 0.0078	0.263 ± 0.136
<i>Umbrina xanti</i>	0.579	2.900
<i>Ophioscion strabo</i>	1.653 ± 1.176	7.515 ± 7.042
<i>Diplectrum pacificum</i>	0.446 ± 0.357	0.560 ± 0.495
<i>Hemicaranx leucurus</i>	0.582 ± 0.539	0.595 ± 0.424
<i>Synodus scituliceps</i>	0.845 ± 0.535	1.512 ± 0.467
<i>Diplectrum macropoma</i>	1.214 ± 0.944	2.598 ± 0.000
<i>Sphyræna ensis</i>	0.701	4.66
<i>Scorpaena sp.</i>	1.725	0.669
<i>Prionotus sp.</i>	0.530 ± 0.298	2.558 ± 0.000
<i>Polydactylus approximans</i>	1.720 ± 0.663	1.171 ± 0.450
<i>Isopisthus remifer</i>	1.776 ± 0.639	2.624 ± 1.933

Table 3

Significant ($p < 0.05$) correlations of Hg concentrations in muscle and liver with weight and total length of bycatch fish.

Species	Correlation	r	Type
<i>Cyclosetta querna</i>	Hg muscle-weight	0.6904	+
	Hg muscle-length	0.6286	+
<i>Diplectrum pacificum</i>	Hg muscle-weight	0.9641	+
	Hg muscle-length	0.9382	+
	Hg liver-weight	0.9743	+
	Hg liver-length	0.9259	+
<i>Hemicaranx leucurus</i>	Hg muscle-weight	0.8967	+
	Hg liver-weight	0.9895	+
<i>Polydactylus approximans</i>	Hg muscle-weight	0.9750	+
<i>Isopisthus remifer</i>	Hg muscle-weight	0.5828	+
<i>Larimus argenteus</i>	Hg muscle-weight	0.3937	–
	Hg muscle-length	0.3898	–
	Hg liver-length	0.3507	–
<i>Haemulopsis axillaris</i>	Hg muscle-weight	0.2880	+
	Hg muscle-length	0.2880	+
<i>Synodus scitulipes</i>	Hg muscle-length	0.9789	+
<i>Diapterus peruvianus</i>	Hg liver-weight	0.9895	+

r—correlation coefficient.

Table 1

Biological information of analyzed ichthyofauna from the SE Gulf of California.

Species	Common name	N	Total weight (g)	Total length (cm)	Feeding habit
<i>Larimus argenteus</i>	Silver drum	67	134 ± 51 (76–366)	23 ± 3 (19–31)	Omnivore, benthic feeder
<i>Haemulopsis axillaris</i>	Yellowstripe grunt	109	234 ± 92 (102–270)	27 ± 3 (24–30)	Carnivore, benthic feeder
<i>Micropogonias ectenes</i>	Slender croaker	31	163 ± 61 (108–466)	27 ± 2 (21–33)	Carnivore, benthic feeder
<i>Selar crumenophthalmus</i>	Bigeye scad	3	130 ± 17 (118–150)	23.5 ± 1 (22.5–25)	Omnivore, benthopelagic feeder
<i>Trachinotus kennedyi</i>	Blackblotch pompano	34	152 ± 38 (78–268)	22 ± 2 (18–27)	Carnivore, benthic feeder
<i>Diapterus peruvianus</i>	Peruvian mojarra	123	149 ± 34 (74–240)	21 ± 2 (16–25)	Carnivore, benthic feeder
<i>Cyclosetta querna</i>	Toothed flounder	10	293 ± 112 (148–446)	29 ± 4 (20.3–33)	Omnivore, benthic feeder
<i>Epinephelus acanthistius</i>	Rooster hind	3	156 ± 21 (134–175)	22 ± 1 (20.6–22.5)	Omnivore, benthopelagic feeder
<i>Umbrina xanti</i>	Polla drum	1	284	28.5	Carnivore, benthic feeder
<i>Ophioscion strabo</i>	Squint-eyed croaker	2	393 ± 140 (294–492)	35 ± 5 (31–38)	Carnivore, benthic feeder
<i>Diplectrum pacificum</i>	Inshore sand perch	5	117 ± 25 (98–148)	21 ± 1 (19.3–22.4)	Carnivore, benthic feeder
<i>Hemicaranx leucurus</i>	Yellowfin jack	3	137 ± 11 (126–148)	25 ± 1 (24–25.5)	Omnivore, benthopelagic feeder
<i>Synodus scituliceps</i>	Shorthead lizardfish	4	380 ± 157 (148–484)	36 ± 6 (28–40)	Carnivore, benthopelagic feeder
<i>Diplectrum macropoma</i>	Mexican sand perch	2	206 ± 74 (154–258)	24 ± 2 (22.5–25.2)	Carnivore, benthic feeder
<i>Sphyræna ensis</i>	Mexican barracuda	1	130	31	Carnivore, pelagic feeder
<i>Scorpaena sp.</i>	Rockfish	1	336	26	Carnivore, benthic feeder
<i>Prionotus sp.</i>	Searobin	2	323 ± 61 (234–350)	30 ± 5 (26–30)	Omnivore, benthic feeder
<i>Polydactylus approximans</i>	Blue bobo	4	149 ± 21 (128–174)	26 ± 2 (23–27)	Omnivore, benthic feeder
<i>Isopisthus remifer</i>	Silver weakfish	12	98 ± 14 (74–128)	57 ± 81 (20.5–24.5)	Omnivore, benthopelagic feeder

Table 4Percentage and species of bycatch that exceeded the maximum permissible limits ($\mu\text{g g}^{-1}$ wet weight) of Hg for human consumption.

Legislation	Limit	(%)	Fish species
Mexico	1.0	0	–
Japan	0.4	26.3	<i>Haemulopsis axillaris</i> , <i>Diapterus peruvianus</i> , <i>Scorpaena</i> sp., <i>Polydactylus approximans</i> , <i>Isopisthus remifer</i>
United Kingdom	0.3	31.6	<i>Haemulopsis axillaris</i> , <i>Diapterus peruvianus</i> , <i>Scorpaena</i> sp., <i>Polydactylus approximans</i> , <i>Isopisthus remifer</i> , <i>Ophioscion strabo</i>

dent's *t* test using GraphPad Prism 4.0 (Graph Pad Software, San Diego, CA).

Biometric information of selected ichthyofauna is presented in Table 1. For this study, nineteen fish species were analyzed. On the basis of the number of specimens, fish species were mainly represented by the Peruvian mojarra *Diapterus peruvianus* (123 individuals), the yellowstripe grunt *Haemulopsis axillaris* (109 individuals), and the silver drum *Larimus argenteus* (67 individuals). In relation to the feeding habits, most of the fish species corresponded to carnivorous (11) followed by omnivorous (8). The feeding guild was benthic or benthopelagic, so all the revised individuals feed, at least partially, from the bottom.

Concentrations of Hg ($\mu\text{g g}^{-1}$ dry weight) in studied ichthyofauna are shown in Table 2. For both tissues, mean Hg concentrations ranged by one order of magnitude. Considering all fish species, mean concentrations in liver ($2.458 \pm 1.997 \mu\text{g g}^{-1}$) were significantly higher ($p < 0.05$) than in muscle ($0.993 \pm 0.670 \mu\text{g g}^{-1}$). With the exception of *Scorpaena* sp. and *Polydactylus approximans*, the sequence of averaged Hg concentrations in analyzed ichthyofauna was liver > muscle. Highest level of Hg in muscle ($2.556 \mu\text{g g}^{-1}$) corresponded to the Peruvian mojarra *D. peruvianus*; in the case of liver, the highest concentration of Hg ($7.515 \mu\text{g g}^{-1}$) occurred in squint-eyed croaker *Ophioscion strabo*. In another study (Ruelas-Inzunza et al., 2008) with some of the fish species used in the current investigation from the SE Gulf of California, it was reported that Hg concentrations in muscle of *H. axillaris* ($1.18 \mu\text{g g}^{-1}$) and *Selar crumenophthalmus* ($0.65 \mu\text{g g}^{-1}$) were comparable to values reported here for the same species, while Hg concentration in *D. peruvianus* ($0.58 \mu\text{g g}^{-1}$) was lower than that reported in the current investigation with the same species. Regarding Hg concentration in similar fish from other regions, average level in muscle ($0.19 \mu\text{g g}^{-1}$) of *S. crumenophthalmus* from SE Asia was comparable to value reported in the current study for the same species; in the case of liver, Hg concentration ($0.15 \mu\text{g g}^{-1}$) in specimens from SE Asia was lower than value reported here (Agusa et al., 2007). In another report from SE Asia, Agusa et al. (2007) measured Hg concentration in muscle ($0.34 \mu\text{g g}^{-1}$) and liver ($0.63 \mu\text{g g}^{-1}$) of *Sphyræna obtusata*; values in muscle were comparable to the level reported here for *Sphyræna ensis* but one order of magnitude lower than Hg concentration in liver. With respect to Hg concentrations in fish species according to the feeding habit, it was found that muscle and liver of carnivores were not significantly ($p > 0.05$) different to corresponding tissues in omnivores. Significant ($p < 0.05$) correlations of Hg levels in muscle and liver with total length and weight of fish are presented in Table 3. In eight of the studied species correlations were positive; i.e. levels of Hg in muscle and liver of fish increased as the organism was longer or heavier.

In relation to legal limits of Hg in fish and fishery products for human consumption established by national and international agencies, percentages of fish species (edible portion) that exceeded such limits are presented in Table 4. It is noticeable that none of the species had levels of Hg in muscle that were equal or above the concentration ($1.0 \mu\text{g g}^{-1}$ wet weight) considered in the Mexican legislation; nevertheless, as legal limits decrease, the number of species over such limits increases. With respect to the legal limit set in the Japanese legislation, 26.3% of the species had Hg levels in the edible portion above $0.4 \mu\text{g g}^{-1}$; if Hg levels in muscle tissue are compared with limits ($0.3 \mu\text{g g}^{-1}$ wet weight) set in the United Kingdom,

31.6% of the muscle samples of bycatch fish species were above that concentration. From a compilation of legal limits of trace metals for diverse fishery products (Nauen, 1983), it is evident that levels set for Hg are variable and they should be taken with caution since some species can have values below some legislation but above another; this is the case of the current study. Additionally, it is worth mentioning that the ratio of methylHg (MeHg) to total Hg in muscle tissue of fish can be as high as 100% (Agah et al., 2007); this issue is of great concern since organic Hg (mainly as MeHg) is more toxic to humans than inorganic forms because of its capacity to pass through biological membranes, high stability and potential of bioaccumulation (Clarkson and Magos, 2006).

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References

- Agah, H., Leermakers, M., Elskens, M., Fatemi, S.M.R., Baeyens, W., 2007. Total mercury and methyl mercury concentrations in fish from the Persian Gulf and the Caspian Sea. *Water Air Soil Pollut.* 181, 95–105.
- Agusa, T., Kunito, T., Sudaryanto, A., Monirith, I., Kan-Antireklap, S., Iwata, H., Ismail, A., Sanguansin, J., Mughtar, M., Tana, T.S., Tanabe, S., 2007. Exposure assessment for trace elements from consumption of marine fish in southeast Asia. *Environ. Pollut.* 145 (3), 766–777.
- Amezcuca-Linares, F., 1990. Los peces demersales de la plataforma continental del Pacífico central de México. Universidad Nacional Autónoma de México, México, Tesis de Doctorado en Ciencias del Mar.
- Cabello, A.M., Martínez, Z., Villegas, L.V., Figuera, B.E., Marcano, L.A., Gómez, A., Vallenilla, O., 2005. Fauna acompañante del camarón como materia prima para la elaboración de productos pesqueros. *Zoot. Trop.* 23 (3), 217–230.
- Clarkson, T.W., Magos, L., 2006. The toxicology of mercury and its chemical compounds. *Crit. Rev. Toxicol.* 36, 609–622.
- Fitzgerald, W.F., Lamborg, C.H., Hammerschmidt, C.R., 2007. Marine biogeochemical cycling of mercury. *Chem. Rev.* 107, 641–662.
- Hylander, L.D., Meili, M., 2003. 500 years of mercury production: global annual inventory by region until 2000 and associated emissions. *Sci. Tot. Environ.* 304, 13–27.
- López-Martínez, J., Morales-Bojórquez, E., Paredes-Mallón, F., Lluch-Belda, D., Cervantes-Valle, C., 2001. La pesquería de camarón de altamar en Sonora. In: Lluch-Belda, D., Elorduy-Garay, J., Lluch-Cota, S., Ponce-Díaz, G. (Eds.), *Centros de Actividad Biológica (BACs) en el Noroeste de México*. CIBNOR-CICIMAR-CONACYT, La Paz, B.C.S., México, pp. 301–312.
- Madrid-Vera, J., Amezcua, F., Morales-Bojórquez, E., 2007. An assessment approach to estimate biomass of fish communities from bycatch data in a tropical shrimp-trawl fishery. *Fish. Res.* 83, 81–89.
- Magalhães, M.C., Costa, V., Menezes, G.M., Pinho, M.R., Santos, R.S., Monteiro, L.R., 2007. Intra- and inter-specific variability in total and methylmercury bioaccumulation by eight marine fish species from the Azores. *Mar. Pollut. Bull.* 54, 1654–1662.
- Moody, J.R., Lindstrom, P.M., 1977. Selection and cleaning of plastic containers for storage of trace element samples. *Anal. Chem.* 49, 2264–2267.
- Mora, C., Robertson, D., 2005. Causes of latitudinal gradients in species richness: a test with fishes of the Tropical Eastern Pacific. *Ecology* 86, 1771–1792.
- Nauen, C.E., 1983. Compilation of legal limits for hazardous substances in fish and fishery products. *FAO Fish. Circ.* 764, 1–102.
- Rábago-Quiroz, C.H., López-Martínez, J., Herrera-Valdivia, E., Nevarez-Martínez, M.O., Rodríguez-Romero, J., 2008. Population dynamics and spatial distribution of flatfish species in shrimp trawl bycatch in the Gulf of California. *Hydrobiologia* 18, 193–202.
- Ruelas-Inzunza, J., Meza-López, G., Páez-Osuna, F., 2008. Mercury in fish that are of dietary importance from the coasts of Sinaloa (SE Gulf of California). *J. Food Comp. Anal.* 21, 211–218.
- UNEP, 1993. Guidelines for monitoring chemical contaminants in the sea using marine organisms. Reference methods for marine pollution studies, No. 6.